



# Investigation of Hepatic Enzymes Changes in Female Teenagers through Sport Tests

Liu Hu<sup>1</sup>, Forough Talaeizadeh<sup>2</sup> and Ahmad Riahi<sup>3\*</sup>

1- Faculty of Physical Education, Beijing University of Technology, China

2- Department of Educational Science and Psychology, Islamic Azad University Marvdasht Branch, Iran

3- Department of Biology, Payam Noor University, Tehran, Iran

Corresponding Author: Ahmad Riahi

Received: 20 September, 2019

Accepted: 01 October, 2019

Published: 19 October, 2019

---

## ABSTRACT

The aim of this study was to determine the effect of eight weeks swimming training on Hepatic Enzymes and Hematological values in young female. Twenty one healthy female were selected by convenience sampling method and were randomly assigned in the two groups: The test (n=15) and control (n=6) groups. The exercise protocol included long-term swimming training lasted for eight weeks and 3 sessions per week and every session lasted for 60 to 90 minutes with intensity of 65-85 percent of maximum heart rate reserve. Blood samples were taken to measure serum hepatic enzymes and hematological values before and after swimming training period. Data were analyzed by parametric (Paired and Independent-Samples t-test) and nonparametric (Wilcoxon and Mann-Whitney U test) for compared within and between groups; and the level of significance was set at  $P < 0.05$ . The level of WBC, RBC, Hb and Hct in exercise group towards the end of period of the training increased significantly. Also, there were no significant differences between groups in the levels of WBC, Hb, PLT, AST and ALT. The variance between group RBC and Hb were significant. Although, the levels of serum AST and ALT levels reduce at the end of the eight weeks swimming training, but these changes did not significantly. The liver function parameters, AST and ALT were decreased after eight weeks swimming training. These findings highlight the importance of imposing restrictions on swimming training and during clinical studies. Future examinations are now essential to clarify the effectiveness of exercise on various parameters.

**Keywords:** Hematological values, Hepatic enzymes, Swimming training.

©2019 GJSR Journal All rights reserved.

---

## INTRODUCTION

The liver is that the main organ for conversion of one chemical species to a different and this interconversion is that the main route for making ready medicine for excretion from the body. The metabolism of medication will result in the formation of chemically reactive intermediates that may play a major role within the induction of hepatic injury. It is necessary that potentially hepatotoxic effects of recent medicine are recognized early throughout drug development. Therefore, in phase I clinical trials, observance of liver perform parameters is necessary. The occurrence of asymptomatic elevations in liver perform tests could be a problem throughout all phases of drug development. An asymptomatic raising of, for instance, liver transaminases throughout clinical trials might be drug related, however other factors, like exercise (Giboney, 2005) and diet (Purkins, et al., 2004), might also have had this effect. A liver is termed "fatty liver" if lipids account for more than five percent of its weight. The mechanisms for the event of liver disease are varied. A decrease within the hepatic oxidation of fatty acids as results of mitochondrial dysfunction can lead to micro vesicular statuses. Another mechanism is association with an imbalance between fat uptake and secretion, with high insulin/glucagon ratio condition cause macro vesicular statuses. Fatty liver are often the consequence of many illness, involving alcohol excess, nonalcoholic steato hepatitis, hepatitis C infection, metabolic disorders, medication effects and nutritional disorders (Laurin et al., 1996).

Most patients with liver disease are asymptomatic, and also the condition is typically discovered due to hepatomegaly or mild abnormalities of serum aminotransferase or alkaline phosphatase levels found on a routine physical examination. One

controlled study demonstrated that a weight reduction program (combined diet and exercise) can improve liver function test results and liver histology in patients with nonalcoholic steato hepatitis. With a weight loss of 4.5 to 6.8 kg, liver transaminase levels often return to normal (Ueno, 1997). Investigators in another study found a correlation between high fat and oil consumption and elevated liver transaminase levels. They concluded that a low-fat diet and exercise could minimize hepatic steatosis (Deems et al., 1994).

Jabbar (2010) Found that resistance training in sedentary males lead to the no significant differences in Aspartate aminotransferase (AST) and Alanine aminotransferase (ALT) activities between creatine training and placebo groups before or after the 8 week training. Therefore, they concluded that eight weeks resistance training period along with creatine monohydrate ingestion does not have adverse effects on these hepatic cellular damage indices. Rezaeeshirazi et al (2011) reported that an eight-week period of aerobic exercise has made no significant changes in the physiological variables such as hepatic enzymes (ALT, AST and ALP). Sreenivasa (2006) in the study on Sixty-five (mean age  $38.7 \pm 9.5$  years; 46 [78%] males) out of 94 patients diagnosed with NASH that participated in their study, found that Moderate intensity aerobic exercise helps in normalizing ALT levels in patients with NASH. Takato (1997) Indicated that restricted diet and exercise therapy such as walking and jogging (for a trial period of 3 months) in twenty-five obese patients with fatty liver, are useful means of improving blood biochemical data and histological findings in liver tissues related to fatty liver.

It is well known that energy-restricted diet and exercise are very useful therapy for obese patients (Mustajoki & Pekkarinen, 2001). These treatments result in the reduction of body weight, blood pressure and serum lipids, and may also improve the fatty liver which accompanies obesity. However, there are few reports demonstrating a relationship swimming exercise and enzymes of liver. In the present study, we investigated the effect of eight weeks swimming training on Hepatic Enzymes and Hematological values in young female.

## 2. Methodology

### 2.1 Subjects

This study was semi-experimental; which compared two groups with pre-test and post-test designs. Furthermore, it plan was confirmed by Research Assembly of Physical Education and Sport Sciences Faculty of Ferdowsi University of Mashhad life save federation. During first stage, the subjects of this study were twenty one healthy female who randomly assigned into the experimental (n=15) and control (n=6) groups. Before starting the program, written informed consents were taken from all subjects. The levels of health and physical activity of the subjects were determined using general practice physical activity questionnaire, physical activity readiness questionnaire and medical survey (including electrocardiogram and blood pressure tests) by a specialist physician (Shephard, 1991). The subjects were nonsmokers, received no drugs and had no metabolic disease and physical impairment affecting their performance. During the second stage, their height was measured in centimeters using a height determiner and their weight was calculated using a digital scale produced by a German company called Beurer (PS07-PS06). The percent of body fat (PBF) was calculated using a body compound determiner (model In-body-720 made in Korea) and based on a method called bioelectrical impedance. All of these measurements were carried out while the volunteers had stopped eating or drinking 4 hours prior to their test, and their bladder, stomach, and bowels were empty.

### 2.2 Exercise protocol

The swimming training program was designed according to in guideline of American College of Sports Medicine (Nelson, 2007) and was performed by professionally qualified physical instructors (an experienced coach) from the health clubs Sports and Health Pioneer East, Mashhad, Iran. Under the supervision of exercise physiologists who assessed and individualized the exercise program for volunteers and supervised their performance during exercise, each participant was trained to achieve a target heart rate reserve (HRR). The exercise protocol included aerobic exercise training lasted for eight weeks and 3 sessions per week and every session lasted for 60 to 90 minutes and with intensity of 65-85 percent of maximum heart rate reserve (MHRR).

According to the MHRR for every single athlete was respectively calculated based on Karvonen equation (1) and was also controlled during exercise by a heart rate monitor (made in Finland–Polar) (Robbert & Landwehr, 2002).

**Equation (1): Target heart rate = [%60 or %70 + [(age-220) - (resting pulse)] + Resting heart rate**

The aerobic training program consisted (20 minutes of warm up exercise, 50 to 60 minutes of aerobic exercise and 10 minutes of cool down exercise) with 65-85 % HRR during each training session. The first to fifth sessions of swimming exercise the HRR was 65% and for the each session the heart rate was added to the end of a swimming exercise sessions. So that in the sessions of sixth to tenth (70%HRR), eleventh to fifteenth sessions (75% HRR), sixteenth to eighteenth sessions (80% HRR) and nineteenth to twenty forth sessions intensity of swimming exercise was maintained in 85% HRR.

### 2.3 Blood sampling

Blood samples in all related studies were collected by venipuncture from forearm vein after at least 15 minutes of sitting at rest or in the supine position. Blood sample were poured into a tube containing K2EDTA and mixed for 15 min before analysis. After centrifuging samples in plastic capillary tubes using Haemato Spin Centrifuge device. Aspartate aminotransferase (AST) and Alanine aminotransferase (ALT) were determined using the Olympus AU400 Chemistry Analyzer.

### 2.4 Statistical analysis

All statistical analyses were performed with SPSS version 15. The average and standard deviation of data were calculated after checking the data distribution normalcy using Kolmogorov-Smirnov test and Homogeneity of variance method. The result of Kolmogorov-Smirnov test in some parameters was not normal. Therefore, data were analyzed by parametric (Paired Samples, Independent-Samples t-test) and nonparametric (Wilcoxon and Mann-Whitney U test) for compared within and between groups; and the level of significance was set at  $P < 0.05$ .

### 3. Results

Table 1. Shows means for age, height, weight and subject's background in sports

Variables	M±SD*
Age (years)	1.53±26.93
Height (cm)	164.33±5.16
Weight (kg)	61.43±165.33

The average, standard deviation and results coming from within the group's changes of levels WBC, RBC, Hb, Hct, PLT, AST of swimmers is presented in table 2. According to the (Table 2), our results show increase in WBC, RBC, Hb and Hct levels in exercise group towards the end of period of the training significantly ( $P < 0.05$ ). Also, there were no significant differences between groups in the levels of WBC, Hb, PLT, AST and ALT. Results showed a variance between group RBC and Hb were significant ( $P < 0.05$ ). The levels of serum AST reduce in exercise group; however, these changes did not significantly ( $P > 0.05$ ).

Table 2. Values of hematological indices and hepatic enzymes before and after eight weeks swimming training (Mean±SD)\*.

Variables	Groups	Pre-test	Post-test	P**	P***
		Mean±SD*	Mean±SD*		
White blood cell count (x 106/mm3)	Exercise group	5706.67±855.62	6742.13±1257.42	0.04*	0.18
	Control group	6150.00±898.33	6133.33±915.78	0.944	
Red blood cell count (x 106/mm3)	Exercise group	4.73±0.29	5.37±0.47	0.00*	0.00*
	Control group	4.83±0.31	4.70±0.20	0.23	
Hemoglobin (gm/dl)	Exercise group	13.90±0.82	14.48±1.22	0.00*	0.17
	Control group	13.38±0.92	13.61±0.81	0.21	
Hematocrit (%)	Exercise group	41.78±2.09	44.27±3.13	0.00*	0.00*
	Control group	41.00±2.49	40.75±1.70	0.64	
Platelets (1000)	Exercise group	259133.42±533.79	267367.71±57908.37	0.37	0.21
	Control group	236500.00±4252.32	225500.00±45715.42	0.07	
AST (U/L)	Exercise group	16.80±5.89	15.40±5.04	0.15	0.28
	Control group	13.17±3.31	13.50±3.72	0.63	

\*Data presented as mean ± standard deviation      \*\* Paired sample t-test      \*\*\* Independent samples t-test

Furthermore, the ALT levels presented that in table 3. Although, the ALT levels reduce at the end of the eight weeks swimming training, but these changes did not significantly ( $P > 0.05$ ).

Table 3. Values of ALT Enzymes before and after eight weeks swimming training (Mean±SD)\*.

Variables	Groups	Pre-test	Post-test	P**		P***	
		Mean±SD*	Mean±SD*	z	p	z	p
ALT (U/L)	Exercise group	19.00±6.38	17.72±4.54	-1.05	0.293	0.161	27
	Control group	15.33±4.08	16.00±3.09	-1.00	0.36		

\*Data presented as mean ± standard deviation      \*\* Wilcoxon      \*\*\* Mann-Whitney U

### 4. Discussion and Conclusion

Changes occur in metabolism depending on the intensity and severity of exercise; changes may be in blood values before and after exercise (Sonmez, 2002). Variations were found among sedentaries and female athletes in some parameters in study done to look at the effects of chronic exercise on some hematological and hepatic enzymes parameters. The results of this study

reveal that the level of WBC increased significantly. The findings from this study are consistent with those reported in the literature. Patlar (2010) reported a significant increase in WBC levels after four weeks of chronic submaximal exercise period. The findings of this study are inconsistent with studies demonstrating that WBC not changed to response exercise. Thus, in studies examining the effects of exercise done on chronic WBC values, Yeh (2006) found that twelve weeks of exercise didn't cause any significant changes in wbc levels. Within the same way, Banfi et al., (2006) reported that WBC levels were similar before and after camp and in study done by Ergun (2006), it couldn't be found a significant increase in WBC levels from blood samples after regular aerobic exercise too. Mashiko (2004) reported a significant decrease in WBC levels of experimental group in camping amount. It is thought that the results of these will increase and reduces was caused by experimental design.

Our study showed significant increased in RBC, Hct and mean hemoglobin concentration after the long swimming training. Halson (2003) reported increase of HGB parameters in trained people and Patlar and Keskin (2007) emphasized decreases and increases in HGB levels after exercise. Yeh (2006) and Umit (2004) haven't found a significant difference at RBC levels of each of the groups after and before exercise program; there are some evidences about exercises done in different intensities that have effects on RBC levels. Once literature was scanned about HCT parameters, Boyali et al., (2006) reported a significant increase at HCT values and Mashiko et al., (2004) identified those twenty days intensive exercise did not cause any changes at HCT levels. Whereas an increase at RBC levels within the different studies, a significant increase could not be found at HGB, HCT and PLT levels (2010). It is thought that these decreases and increases were caused by exercise protocols that applied the sector-specific. Alterations of the hematological variables can influence physical performance. Also, Performance in exercise depends on each the amount of hemoglobin and therefore the development of ability of carrying oxygen (Gleeson, 2003).

Intensive exercise can cause significant differences in hemoglobin values (Nieman & Pedersen, 1999). Oxygen from red blood cells in tissues is connected to hemoglobin which provides the oxygen to active tissues. Organism's oxygen demand increases throughout training. In parallel to the present increase, the circulatory and respiratory systems should show a physiological adaptation. Then the oxygen would like of tissues and therefore the amount of oxygen of the cardiovascular system increases (Ghosh, 1985). Especially, evaluation of blood volume in young is complex. There are conflicting results in literature regarding changes in blood volume per unit body mass increases with age. Women have lower hemoglobin concentration values and it would lead to reduced oxygen carrying capacity (Armstrong et al., 2001). In fact, exercise intensity and duration are two necessary components in exercise or regular training. Therefore, it is potential that duration and intensity of this training protocol were enough to have an effect on hematological variables.

Our study showed declines in AST and ALT concentration after the long-term swimming training. This finding was supported by Sreenivasa et al., (2006) and Davoodi et al., (2012). Davoodi et al., (2012) in their study found that eight weeks selected aerobic exercise lead to the reduces in serum AST and ALT in experimental group rather than control group. The findings from the present study are inconsistent with those reported in the literature. Nie et al., (2011) found statistically a significant increase at AST and ALT levels after exercise, also in other study, it was reported a significant increase at AST and ALT values of athletes running ultra marathon after and before competition (Wu et al., 2004). In excessive muscle forced exercise-induced, AST and ALT levels in blood can raise in muscle damages. Membrane permeability, changes which cause these enzymes to leak into serum, occur related to muscle damage exercise-induced (Brancaccio et al., 2010). Furthermore, Aerobic activity stimulates lipid oxidization and hinders lipogenesis inside the liver (Lisa, 2011; Ruderman, 2003). This is often mediate by the activation of AMPK pathway. This enzyme is activated when the ratio of AMP to ATP in tissues as in an aerobics. Studies show that a reduced or absent activity of hepatic SCD-1 is crucial to the activation of AMPK. Recent animal studies have found a significant reduction of SCD-1 activity after aerobic exercise (Ruderman, 2003). AMPK remains activated after the completion of aerobic activity in liver, adipose tissue and muscle. In liver it reduces lipogenesis through direct inactivation of ACC enzyme and activation of MCD. It also reduces related gene expressions of lipogenic enzymes; ACC and FAS. Activation of MCD in turn reduces the amount of Malonyl-CoA that is an inhibitor of CPT-1. The latter enzyme regulates fatty acid transfer to the mitochondria and thus stimulates hepatic lipid oxidation (Ruderman et al., 2003).

As a result, aerobics training can cause increased insulin sensitivity and hepatic lipid oxidization and also reduced activity and inhibition of lipogenic enzymes. All of them contribute to a reduction of hepatic fat (Scott Rector et al., 2008). This can explain the significant reduction of blood serum ALT concentration of patients within the group with added aerobic training.

After eight swimming training, hematologic parameters showed differences when compared to resting values. Besides, there have been no significant variations in ALT and AST levels after eight weeks swimming training. As a result, several changes in metabolism occur after long-term exercise. Several other factors play a major role, such as adaptation to exercise, adaptation of cardiovascular, physical and physiological balance in hematological levels. It may be said that more comprehensive studies on examine training should be done to determine the relation among hematologic and hepatic enzymes values and sportive performance.

## REFERENCES

1. Armstrong N, Welsman J, 2001. Peak oxygen uptake in relation to growth and maturation in 11-to-17-year-old humans. *Eur. J. Appl. Physiol.* 85:546-551.
2. Banfi G, Del-Fabro M, Mauri C, Corsi M, Melegati G, 2006. Hematological Parameters In Higly Elite Rugby Players During A Competitive Season. *Clin. Lab. Haematol.* 28(3):183-198.
3. Boyali E, Cakmakçı O, Patlar S Cakmakçı E 2006. Effects Of Camp Term On Some Hematological Parameters In Male Taekwondoers. *Uluslararası Spor Bilimleri Kongresi bildiri kitabı, Mugla.* 9: 280-281.
4. Brancaccio P, Lippi G, Maffulli N, 2010. Biochemical markers of muscular damage. *Clin. Chem. Lab. Med.* 48(6): 757-767.
5. Davoodi M, Moosavi H, Nikbakht M, 2012. The effect of eight weeks selected aerobic exercise on liver parenchyma and liver enzymes (AST, ALT) of fat liver patients. *J Shahrekord Univ Med Sci.* 14(1): 84-90.
6. Deems R, Friedman L, Friedman M, Munoz S, Deems D, Maddrey W, 1994. Relationship between liver biochemical tests and dietary intake in patients with liver disease. *J Clin Gastroenterol.* 18: 304-308.
7. Ergun M, Tengiz I, Turk U, Senisik S, Alioglu E, Yuksel O, 2006. The Effect of Long Term Regular Exercise On Endothelial Functions, Inflammatory And Thrombotic Activity In Middle Aged. Healthy Men. *J. Sport Sci. Med.* 5: 266-275.
8. Ghosh , A, Ahuja A, Khanna G, 1985. Pulmonary capacities of different groups of sportmen in India. *Br. J. Sports Med.* 19: 232-234.
9. Giboney P, 2005. Mildly elevated liver transaminase levels in the asymptomatic patient. *Am Fam Physician.* 71: 1105-1110.
10. Gleeson M, 2003. Immunological Responses to Overreaching in Cyclists. *Med. Sci. Sports Exerc.* 35(5): 854-861.
11. Halson S, Lnacaster G, Jeukendrup A, Gleeson M, 2003. Immunological Responses to Overreaching in Cyclists. *Med. Sci. Sports Exerc.* 35(5): 854-861.
12. Jabbar B, Gaeini A, Nikbakht H, Farzad Z, Hamdollah H, Mahdi B, 2010. The Effect of Creatine Monohydrate Supplementation with Resistance Training on Liver Responses in Sedentary Males. *World Applied Sciences Journal.* 10(3): 316-321.
13. Laurin J, Lindor K, Crippin J, Gossard A, Gores G, Ludwig J, 1996. Ursodeoxycholic acid or clofibrate in the treatment of non-alcohol-induced steatohepatitis: a pilot study. *Hepatology* 23: 1464-1467.
14. Lisa B, Wagner V, Mary E, 2011. Rinella The role of insulin-sensitizing agents in the treatment of nonalcoholic steatohepatitis. *Ther Adv Gastroenterol.* 4(4): 249-263.
15. Mashiko T, Umeda T, Nakaji S, Sugawara K, 2004. Effects Of Exercise On The Physical Condition Ofcollege Rugby Players During Summer Training Camp. *Br. J. Sports Med.* 38: 186-190.
16. Mustajoki P, Pekkarinen T 2001. Very low energy diets in the treatment of obesity. *obesity reviews.* 2: 61-72.
17. Nelson M, Rejeski W, Blair S, Duncan P, Judge J, King A, 2007. Physical activity and public health in older adults: recommendation from the American College of Sports Medicine and the American Heart Association. *Med Sci Sports Exerc.* 39: 1435-1445.
18. Nie J, Tong T, George K, Fu F, Lin H, Shi Q, 2011 . Resting and post-exercise serum biomarkers of cardiac and skeletal muscle damage in adolescent runners. *Scand J Med Sci Sports.* 21(5): 625-629.
19. Nieman D, Pedersen B, 1999. Exercise and Immune Function: Recent Development. *Sports Med.* 27: 73-80.
20. Patlar S, 2010. Effects of acute and 4-week submaximal exercise on leukocyte and leukocyte subgroups. *Isokinetics Exercise Sci.* 18: 145-148.
21. Patlar S, Keskin E, 2007. The Effects of Glycerol Supplement on Various Hematologic Parameters in Sedentaries and The Athletes who Exercise Regularly. *Exercise.* 1: 23-35.
22. Purkins L, Love E, Eve M, Wooldridge C, Cowan C, Smart T, 2004. The influence of diet upon liver function tests and serum lipids in healthy male volunteers resident in a Phase I unit. *Br J Clin Pharmacol.* 57: 199-208.
23. Rezaeeshirazi R, Hossini F, Tarasi Z, Shaygan Asl N, 2011. The Effect of an Aerobic Exercise Program on General Health and Hepatic Enzymes Among Incarcerated Addicts. *Australian Journal of Basic and Applied Sciences.* 5(10): 1191-1194.
24. Robbert A, Landwehr R, 2002. The supporting history of the “HRmax=220-age” equation. *Journal of Exercise Physiology online.* 5(2): 1-10.
25. Ruderman N, Park H, Kaushik V, Dean D, Constant S, Prentki M, 2003. AMPK as a metabolic switch in rat muscle, liver and adipose tissue after exercise. *Acta Physiol Scand.* 178: 435-442.
26. Scott Rector R, Thyfault J, Tyler Morris R, Laye M, Borengasser S, Booth F, 2008. Daily exercise increases hepatic fatty acid oxidation and prevents steatosis in Otsuka Long-Evans Tokushima Fatty rats. *Am J Physiol Gastrointest Liver Physiol.* 294: G619-G626.
27. Shephard R, 1991. Readiness for Physical Activity. *Sports Medicine* 1:359.
28. Sonmez G, 2002. Exercise and sports physiology. 1 ed. Bolu: Ata Publications.
29. Sreenivasa Baba C, Alexander G, Kalyani B, Pandey R, Rastogi S, Pandey A, 2006. Effect of exercise and dietary modification on serum aminotransferase levels in patients with nonalcoholic steatohepatitis. *Journal of Gastroenterology and Hepatology.* 2: 191-198.
30. Takato U, Sugawara H, Sujaku K, Hashimoto O, Tsuji R, Tamaki S, 1997. Therapeutic effects of restricted diet and exercise in obese patients with fatty liver. *Journal of Hepatology.* 21: 103-107.
31. Ueno T, Sugawara H, Sujaku K, Hashimoto O, Tsuji R, Tamaki S, 1997. Therapeutic effects of restricted diet and exercise in obese patients with fatty liver. *J Hepatol.* 27:103-107.
32. Umit K, Yalcin O, Gunduz F, Kuru O, Herbert J, Baskurt O, 2004. Effect Of Antioxidant Vitamin Treatment On The Time Course Of Hematological And Hemorheological Alterations After An Exhausting Exercise Episode In Human Subjects. *Appl. Physiol.* 98: 1272-1279.
33. Wu H, Chen K, Shee B, Huang Y, Yang R, 2004. Effects of 24 h ultra-marathon on biochemical and hematological parameters. *World J. Gastroenterol.* 10(18): 2711-2714.

34. Yeh SH, Chuang H, Lin LW, Hsiao CY, Eng H. 2006. Regular Tai Chi Chuan Exercise Enhances Functional Mobility And Cd4cd25 Regulatory T Cells. *British J. Sports Med.* 40: 239-243.
35. Ahmadipour, A., Shaibani, P., & Mostafavi, S. A. (2019). Assessment of empirical methods for estimating potential evapotranspiration in Zabol Synoptic Station by REF-ET model. *MedBioTech Journal*, 03(01), 1-4. doi:10.22034/mbt.2019.80823
36. Aida Badamchi Shabestari, B. A. A., Maryam Shekarchi, Seyed Mojtaba Mostafavi. (2018). Development of Environmental Analysis for Determination of Total Mercury in Fish Oil Pearls by Microwave Closed Vessels Digestion Coupled with ICP-OES. *Ekoloji*, 27(106), 1935 .
37. Aida Badamchi Shabestari, S. M. M., Hanieh Malekzadeh. (2019). Force Degradation Comparative Study on Biosimilar Adalimumab and Humira. *Revista Latinoamericana de Hipertensión*, 13(06), 496-509 .
38. Al Tariq, Z. (2018). Clinical, Biochemical and Immunological Profiles of HIV Patients Developing Immune Reconstitution Inflammatory Syndrome (IRIS). *MedBioTech Journal*, 02(01), 21-28. doi:10.22034/MBT.2018.61528
39. Amanlou, M., & Mostafavi, S. M. (2017). In silico screening to aim computational efficient inhibitors of caspase-9 by ligand-based pharmacophore modeling. *MedBioTech Journal*, 01(01), 34-41. doi:10.22034/mbt.2017.60337
40. Amir Yaghoubi Nezhad, S. H., Atefeh Mehrabi Far, Masoumeh Piryaee, Seyed Mojtaba Mostafavi. (2019). Investigation of Shigella Lipopolysaccharides Effects on Immunity Stimulation of Host Cells. *International Transaction Journal of Engineering, Management, Applied Sciences and Technologies*, 10, 465 .
41. Banaei, M. (2019). Investigating the effect of hyperglycemia on iron, copper, and zinc micronutrients in diabetic patients. *MedBioTech Journal*, 03(01), 13-15. doi:10.22034/mbt.2019.80827
42. Basse, U., & Edoamodu, O. (2018). In vivo Investigation of Haematological and Histological Effects of Leaves Extracted from some Herbs on *Plasmodium berghei*. *MedBioTech Journal*, 02(01), 35-41. doi:10.22034/MBT.2018.61608
43. Bobadilla, C., & Roja, R. (2018). Antioxidant and Chemical Activity of South American Chocolate. *MedBioTech Journal*, 02(01), 29-34. doi:10.22034/MBT.2018.61529
44. Bouaffad, H., & Hayyani, N. (2018). Determination of Elemental Impurities Ciprofloxacin hydrochloride and Fluconazole via ICP-OES. *MedBioTech Journal*, 02(01), 41-46. doi:10.22034/MBT.2018.61612
45. Emami, M. (2019). Evaluation of the TPS gene as an internal control gene in *Gossypium* genotype for the quantitative detection of cotton transgenic product using Time-Real PCR technique. *MedBioTech Journal*, 03(01), 20-21. doi:10.22034/mbt.2019.80836
46. Farah, T., & Kalsoum, C. (2018). Analysis of E.Coli from Waste Effluents of a Farm Business. *MedBioTech Journal*, 02(03), 85-86. doi:10.22034/MBT.2018.76929
47. Haghghi, Z., & Asadi, M. (2019). The Effects of Chitosan-based nanofibers /PEO/ henna extract on recovery of superficial second-degree burn in rat. *MedBioTech Journal*, 03(01), 26-28. doi:10.22034/mbt.2019.80840
48. Heidari, S., Imani, M., & Mostafavi, S. M. (2017). A Validated and Rapid HPLC Method for Quantification of Human Serum Albumin in Interferon beta-1a Biopharmaceutical Formulation. *MedBioTech Journal*, 1(01), 29 .
49. Jafari, S., & Mostafavi, S. A. (2019). Investigation of nitrogen contamination of important subterranean water in the plain. *MedBioTech Journal*, 03(01), 10-12. doi:10.22034/mbt.2019.80826
50. Jagat, T., & Mahesh, S. (2018). Evaluation of Nutritional Values of Some Meals Containing Soybean Oil. *MedBioTech Journal*, 02(02), 69-75. doi:10.22034/MBT.2018.76926
51. Jeon, K. L., & Hu, P. (2017). Determination of Antioxidant Effects of Some Medicinal Plants and Investigation of their Physicochemical Characters. *MedBioTech Journal*, 01(02), 65-72. doi:10.22034/MBT.2017.60370
52. Katra, R., & Lupetki, J. (2018). The Effect of Weeds on Cropping System for Sustaining Food Security. *MedBioTech Journal*, 02(02), 50-53. doi:10.22034/MBT.2018.76915
53. Koko, A., & Yako, L. (2018). How Nutritional Composition of Commonly Consumed Vegetable Changes under the Influence of Fermentation. *MedBioTech Journal*, 02(02), 59-64. doi:10.22034/MBT.2018.76923
54. Krishitiana, N., & Kimatova, L. (2018). Antibacterial Study and Green Preparation of Silver Nanoparticles through Some Plants. *MedBioTech Journal*, 02(02), 54-58. doi:10.22034/MBT.2018.76920
55. Laksian, R., & Rezaei, M. (2019). Evaluation of modified montmorillonite clay with magnetite nanoparticles. *MedBioTech Journal*, 03(01), 5-9. doi:10.22034/mbt.2019.80824
56. Mostafavi, S. M. (2015). 3D Graphene Biocatalysts for Development of Enzymatic Biofuel Cells: A Short Review. *Journal of NanoAnalysis*, 2(2), 57-62 .
57. Mostafavi, S. M., Bagherzadeh, K., & Amanlou, M. (2017). A new attempt to introduce efficient inhibitors for Caspas-9 according to structure-based Pharmacophore Screening strategy and Molecular Dynamics Simulations. *MedBioTech Journal*, 01(01), 1-8. doi:10.22034/mbt.2017.60325
58. Mostafavi, S. M., Eissazadeh, S., & Piryaee, M. (2019). Comparison of Polymer and Ceramic Membrane in the Separation of Proteins in Aqueous Solution Through Liquid Chromatography. *Journal of Computational and Theoretical Nanoscience*, 16(1), 157-164 .
59. Mostafavi, S. M., Malekzadeh, H., & Taskhiri, M. S. (2019). In Silico Prediction of Gas Chromatographic Retention Time of Some Organic Compounds on the Modified Carbon Nanotube Capillary Column. *J. Comput. Theor. Nanosci*, 16, 151-156 .
60. Muhammad, O., & Lama, R. (2018). The Effect of Diclofenac Sodium on Blood Vessel Formation. *MedBioTech Journal*, 02(02), 76-81. doi:10.22034/MBT.2018.76927
61. Parvanian, S., Mostafavi, S. M., & Aghashiri, M. (2016). Multifunctional Nanoparticle Developments in Cancer Diagnosis and Treatment. *Sensing and Bio-Sensing Research*, 1(2), 22 .
62. Pasban, A., Mostafavi, S. M., Malekzadeh, H., & Mohammad Nazari, B. (2017). Quantitative Determination of LPG Hydrocarbons by Modified Packed Column Adsorbent of Gas Chromatography Via Full Factorial Design. *Journal of NanoAnalysis*, 4(1), 31-40 .

63. Rahman, Z., & Vaheed, S. (2018). Colonization and Optimization of Some Fungal Mycelium through Metal Biosorbent. *MedBioTech Journal*, 02(03), 103-107. doi:10.22034/MBT.2018.76933
64. Samira Eissazadeh, M. P., Mohammad Sadegh Taskhiri, & Mostafavi, S. M. (2019). Improvement of Sensitivity of Antigen-Antibody Detection of Semen Through Gold Nanoparticle. *Research Journal of Pharmaceutical, Biological and Chemical Sciences*, 10(1), 144 .
65. Samira Eissazadeh, M. P., & Mostafavi, S. M. (2019). Measurement of Some Amino Acid Using Biosensors Based on Silicon-Based Carbon Nanotubes. *Journal of Computational and Theoretical Nanoscience*, 16, 1 .
66. Samira Eissazadeh, S. M. M., Masoumeh Piryaeei, & Taskhiri, M. S. (2019). Application Of Polyaniline Nanostructure Based Biosensor For Glucose And Cholesterol Detection. *Research Journal of Pharmaceutical, Biological and Chemical Sciences*, 10(1), 150 .
67. Seydoglu, S. (2017). Grafting of Aryl Group over Carbon Electrode Surface and Investigation of its Stability. *MedBioTech Journal*, 01(01), 48-51. doi:10.22034/MBT.2017.60339
68. Seyed Mojtaba Mostafavi, H. M., & Taskhiri, M. S. (2019). In Silico Prediction of Gas Chromatographic Retention Time of Some Organic Compounds on the Modified Carbon Nanotube Capillary Column. *Journal of Computational and Theoretical Nanoscience*, 16(151), 156 .
69. Seyed Mojtaba Mostafavi, A. R., Mina Adibi, Farshid Pashae, Masoumeh Piryaeei. (2011a). Electrochemical Investigation of Thiophene on Glassy Carbon Electrode and Quantitative Determination of it in Simulated Oil Solution by Differential Pulse Voltammetry and Amperometry Techniques. *Asian Journal of Chemistry*, 23(12), 5356-5360 .
70. Seyed Mojtaba Mostafavi, A. R., Mina Adibi, Farshid Pashae, Masoumeh Piryaeei. (2011b). Modification of Glassy Carbon Electrode by a Simple, Inexpensive and Fast Method Using an Ionic Liquid Based on Imidazolium as Working Electrode in Electrochemical Determination of Some Biological Compounds. *Asian Journal of Chemistry*, 23 .(12)
71. Seyed Mojtaba Mostafavi, M. P., Ahmad Rouhollahi, & Mohajeri, A. (2014a). Separation and Quantification of Hydrocarbons of LPG Using Novel MWCNT-Silica Gel Nanocomposite as Packed Column Adsorbent of Gas Chromatography. *Journal of NanoAnalysis*, 1(01), 01 .
72. Seyed Mojtaba Mostafavi, M. P., Ahmad Rouhollahi, & Mohajeri, A. (2014b). Separation of Aromatic and Alcoholic Mixtures using Novel MWCNT-Silica Gel Nanocomposite as an Adsorbent in Gas Chromatography. *Journal of NanoAnalysis*, 1(01), 11 .
73. Shamsipur, M., Beigi, A. A. M., Teymouri, M., Poursaberi, T., Mostafavi, S. M., Soleimani, P., . . . Tash, S. A. (2012). Biotransformation of methyl tert-butyl ether by human cytochrome P450 2A6. *Biodegradation*, 23(2), 311-318 .
74. Tasnim, T., & Farasat, A. (2018). The Bioproduction of Ethanol through Isolation of Some Local Bacteria. *MedBioTech Journal*, 02(03), 132-135. doi:10.22034/mbt.2018.80815
75. Z. Man, A. G. E., S. M. Mostafavi, & Surendar, A. (2019). Fuel oil characteristics and applications: economic and technological aspects. *Petroleum Science and Technology* .